

Phenotypic detection of inducible clindamycin resistance by D-test in *Staphylococci* from various clinical samples



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Abstract:

Background: *Staphylococcus aureus* is a major opportunistic human pathogen causing many clinical infections. Three MLSB phenotypes are known in *S. aureus*, a constitutive resistant phenotype (cMLSB), a clindamycin-susceptible phenotype in vitro with inducible resistance in vivo (iMLSB), and a clindamycin-susceptible and macrolide-streptogramin B-resistant phenotype (MSB). **Method:** This prospective-observational study was carried out in Department of Microbiology, NIMS medical college, Jaipur, Rajasthan from July 2023 to Sept 2024. A total of 50 *Staphylococci* were isolated from different types of samples. All the *Staphylococcus aureus* isolates that were resistant to erythromycin were tested for inducible Clindamycin resistance by double disk approximation test (D-test) as per CLSI guidelines. **Result:** A total of 50 *Staphylococci* isolates, MSSA were 18 (36%) followed by CONS 12 (24%), MR-CONS 12 (24%) and MRSA 8 (16%). Out of total 50 isolates, 17 (34%) showed inducible resistant by D-test. In which MSSA were 3 (18%), CONS 5 (29%), MR-CONS 4 (24%) and MRSA 5 (29%). Inducible resistance was observed among various clinical samples like, Blood samples 10 (60%) followed by Pus 5 (30%), Wound Swab 1(5%), and High Vaginal Swab 1(5%). **Discussion:** In this study we found 34% inducible clindamycin resistance by D-test. Similar findings observed in various studies also. **Conclusion:** Highest percentage of inducible resistance found in isolates from Blood sample. Clinical microbiology laboratories should report inducible Clindamycin resistance in *Staphylococcus aureus* and D-test can be used as a simple, auxiliary and reliable method to Delineate inducible and constitutive Clindamycin resistance in routine clinical laboratories.

Keywords: MLSB, D-test, *ermA*, *msrA*.

Introduction:

Staphylococcus aureus is a major opportunistic human pathogen causing many clinical infections. The rates of infections caused by *staphylococci*, both community, and hospital-acquired strains, are increasing steadily.[1] Erythromycin and clindamycin are used as topical therapeutic agents for treatment of skin and soft tissue infections associated with *Staphylococcal* infections [2].

Three MLSB phenotypes are known in *S. aureus*, a constitutive resistant phenotype (cMLSB), a clindamycin-susceptible phenotype in vitro with inducible resistance in vivo (iMLSB), and a clindamycin-susceptible and macrolide-streptogramin B-resistant phenotype (MSB).

Clindamycin, a macrolide-lincosamide streptogramin B (MLSB) family of antibiotics, is the drug usually advocated for treating *Staphylococcal* infection.[3] Resistance to MLSB arises mainly by an active efflux mechanism coded by the *msrA* gene or *ermA* gene. In inducible resistance, rRNA methylase is produced only in the presence of an inducing agent which can be any of the antibiotics in the MLSB family such as erythromycin [4,5]. Isolates with only inducible resistance are resistant to erythromycin but appear susceptible to clindamycin in vitro [6-8]. The purpose of this study is to detect inducible clindamycin resistance and antibiotic susceptibility

pattern among *staphylococcus aureus* isolates in various clinical specimens.

Materials and methods:

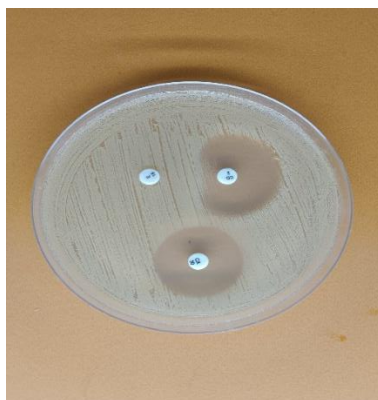
This prospective-observational study was carried out in Department of Microbiology, NIMS medical college, Jaipur, Rajasthan from July 2023 to Sept 2024 after approval of ethical committee. The study population included patients of all age groups and sex visiting the inpatient and outpatient departments of the hospital to whom culture tests had been referred.

A total of 50 *Staphylococci* were isolated from different types of samples; Pus (n=10), Blood (n=31), CSF (n=1), wound swab (n=4), high vaginal swab (n=2), ET (n=1), and nasal swab (n=1) received in the bacteriology lab of NIMS hospital, using standard microbiological techniques. [9, 10].

Antibiotic susceptibility testing was done on Muller Hinton agar plate as per standard guidelines and procedure. All the *Staphylococcus aureus* isolates that were resistant to erythromycin were tested for inducible Clindamycin resistance by double disk approximation test (D-test) as per CLSI guidelines. In this test, turbidity of *Staphylococcus aureus* is compared with 0.5% McFarland's standard. The suspension was lawned over the MHA plate and the antibiotic disc was placed over the lawned plated. An erythromycin disk (15µg) and Clindamycin (2µg) were placed 15mm apart edge-to-edge on MHA plate. Methicillin resistance was identified by placing

Cefoxitin (30µg) disc on MHA plate and interpreted as per CLSI guidelines. Plates were analysed after overnight of incubation at 37°C. On next day the

plates were examined for the zone of inhibition around the antibiotic disc. The diameter of the inhibitory zone includes the diameter of the disc.



Result: A total of 50 *Staphylococci* isolates, MSSA were 18 (36%) followed by CONS 12 (24%), MR-CONS 12 (24%) and MRSA 8 (16%). Out of total 50 isolates, 17 (34%) showed inducible resistant by D-test. In which MSSA were 3 (18%), CONS 5 (29%),

MR-CONS 4 (24%) and MRSA 5 (29%). Inducible resistance was observed among various clinical samples like, Blood samples 10 (60%) followed by Pus 5 (30%), Wound Swab 1(5%), and High Vaginal Swab 1(5%).

Table 1: Gender wise distribution of clinical Isolates

Gender	No of Isolates
Male	35 (70%)
Female	15 (30%)
Total	50 (100%)

Figure I: Gender wise distribution of clinical Isolates

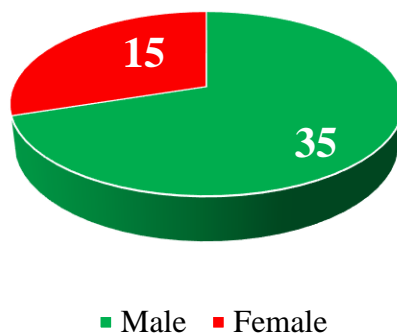
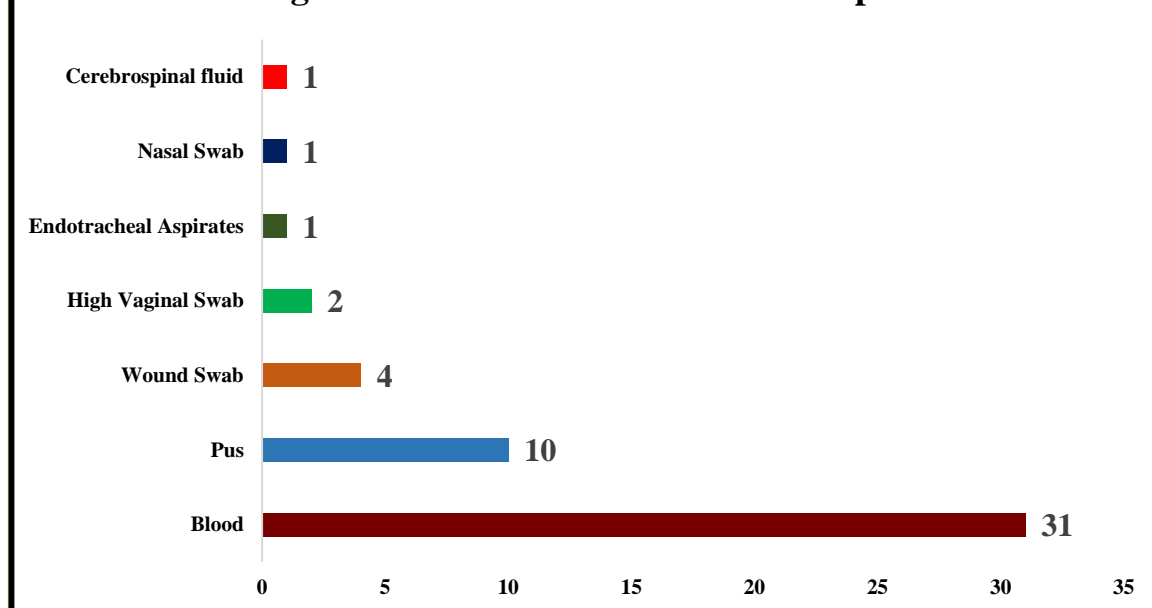
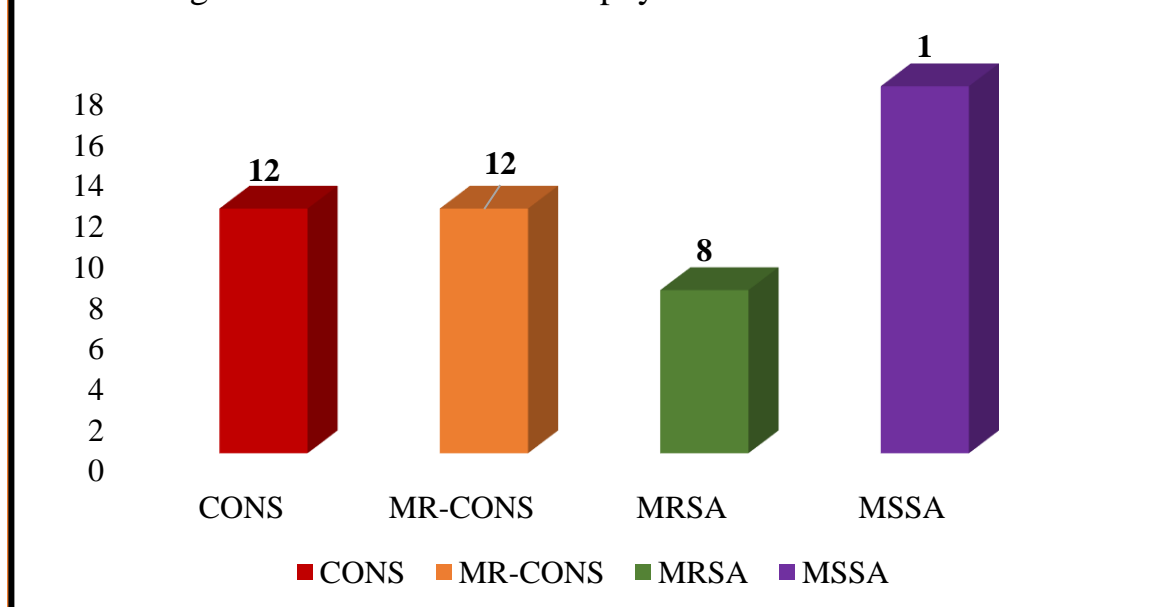


Table 2: Distribution of clinical samples according to D-test positive

Type of samples	No. of samples	D-Test Positive
Blood	31 (62%)	10 (60%)
Pus	10 (20%)	05 (30%)
Wound Swab	04 (8%)	01 (5%)
High vaginal swab	02 (4%)	01 (5%)
Endotracheal aspirates	01 (2%)	00
Nasal swab	01 (2%)	00
Cerebrospinal fluid	01 (2%)	00
Total	50 (100%)	17 (34%)

Figure II: Distribution of clinical samples

Organisms	Number of isolates	D-test Positive
Coagulase negative staphylococcus spp. (CONS)	12 (24%)	05 (29%)
Methicillin resistance coagulase negative staphylococcus spp. (MR-CONS)	12 (24%)	04 (24%)
Methicillin resistance staphylococcus aureus (MRSA)	08 (16%)	05 (29%)
Methicillin sensitive staphylococcus aureus (MSSA)	18 (36%)	03 (18%)
Total	50	17

Figure III: Distribution of Staphylococci Clinical Isolates

Discussion: In this study we found 34% inducible clindamycin resistance by D-test. Shantala G B et al in their study observed that 57 (24.89%) had the inducible clindamycin resistance. (11)

Conclusion: Highest percentage of inducible resistance found in isolates from Blood sample.

Clinical microbiology laboratories should report inducible Clindamycin resistance in Staphylococcus aureus and D-test can be used as a simple, auxiliary and reliable method to Delineate inducible and constitutive Clindamycin resistance in routine clinical laboratories.

Bibliography:

1. Adhikari RP, Shrestha S, Barakoti A, Amatya R. Inducible clindamycin and methicillin resistant *Staphylococcus aureus* in a tertiary care hospital, Kathmandu, Nepal. BMC Infect Dis. 2017;17(1):483. <https://doi.org/10.1186/s12879-017-2584-5>.
2. Ajantha GS, Kulkarni RD, Shetty J, Shubhada C, Jain P. Phenotypic detection of inducible clindamycin resistance among *Staphylococcus aureus* isolates by using the lower limit of recommended inter-disk distance. Indian J Pathol Microbiol 2008; 51(3):376- 378.
3. Prabhu K, Rao S, Rao V. Inducible clindamycin resistance in *Staphylococcus aureus* isolated from clinical samples. J Lab Physicians. 2011;3(1):25-7.
4. Fiebelkorn KR, Crawford SA, McElmed MI, et al. Practical disk diffusion method for detection of inducible clindamycin resistance in *Staphylococcus aureus* and coagulase negative staphylococci. J Clin Microbiol. 2003; 41: 4740-4.
5. Leclercq R. Mechanisms of resistance to macrolides and lincosamides: nature of the resistance elements and their clinical implications. Clin Infect Dis. 2002;34(4):482-92.
6. Drinkovic D, Fuller ER, Shore KP, et al. Clindamycin treatment of *Staphylococcus aureus* expressing inducible clindamycin resistance. J Antimicrob Chemother. 2011; 48:315-6.
7. Fiebelkorn KR, Crawford SA, McElmed MI, et al. Practical disk diffusion method for detection of inducible clindamycin resistance in *Staphylococcus aureus* and coagulase negative staphylococci. J Clin Microbiol. 2003; 41:4740-4.
8. Gadepalli R, Dhawan B, Mohanty S, et al. Inducible clindamycin resistance in clinical isolates of *Staphylococcus aureus*. Indian J Med Res. 2006; 123:571-3.
9. Marin LFC, Arciniegas GE, Vivas MC. Characterization of *Staphylococcus aureus* isolates that colonize medical students in a hospital of the city of Cali, Colombia. Int J Microbiol. 2015;2015:358489. <https://doi.org/10.1155/2015/358489>.
10. Safarpour Dehkordi F, Gandomi H, Basti AA, Misaghi A, Rahimi E. Phenotypic and genotypic characterization of antibiotic resistance of methicillin-resistant *Staphylococcus aureus* isolated from hospital food. Antimicrob Resist Infect Control. 2017;6(1):1-1. <https://doi.org/10.1186/s13756-017-0257-1>
11. Shantala G B, Adithi S Shetty, Rahul Rao K, Vasudeva and Nagarathnamma T ,Detection of inducible Clindamycin resistance in clinical isolates of *Staphylococcus aureus* by the Disc Diffusion Induction Test , Journal of Clinical and Diagnostic Research. 2011 Feb, Vol-5(1):35-37.